

1. Principle:

This method involves the use of HS-GC-MS (single quad) for the determination of N-Nitrosodimethylamine (NDMA).

2. Scope:

This method is applicable to Active Pharmaceutical Ingredients (API) and Powdered Tablets. These samples are preliminarily screened in Scan mode, **Procedure A**. If NDMA is found to be present, samples undergo a NDMA Quantification method, outlined in **Procedure B**. If no NDMA is found samples follow **Procedure C**.

3. Apparatus:

- 3.1 **HS-GC-MS:** Shimadzu GC-2010Plus Gas Chromatograph (Item No. 1043B), Headspace HS-20 auto-sampler (Item No. 1043A), GC-MS QP2020 Mass Spectrometer (Item No. 1043C), or equivalent.
- 3.2 **Autopipettes, Gilson or equivalent:** (1,000 μL , 200 μL , 100 μL , 20 μL , 10 μL).
- 3.3 **Capillary GC Column:** Restek Rtx-624 (30m x 0.25 mm I.D, 1.4mm).

4. Reagents:

- 4.1 **Solvent:** Dimethyl sulfoxide (DMSO), (Merck SupraSolv, Dimethyl Sulfoxide for headspace gas chromatography), Catalogue no. 101900.
- 4.2 Helium gas (CP grade), BOC supplied or equivalent.
- 4.3 **Reference Standard:** NDMA (N-nitrosodimethyl amine), Restek Catalog no. 31427, Concentration **1000 $\mu\text{g}/\text{mL}$** in Methanol (Reference Standard).
- 4.4 **Quality Control Standard:** NDMA (N-nitrosodimethyl amine), Supelco Catalog no. 48670, Concentration **200 $\mu\text{g}/\text{mL}$** in Methanol (QC).

Prepared solutions are protected from light prior to analysis.

5. Calibration Standards:

5.1 Stock standard Solution (NDMA) 200 $\mu\text{g}/\text{mL}$

Dilute 200 μL of NDMA Reference Standard (4.3) to 1mL in DMSO, in a volumetric flask.

5.2 1 $\mu\text{g}/\text{mL}$ NDMA Standard:

Dilute 10 μL of stock standard (200 $\mu\text{g}/\text{mL}$) (5.1) to 2mL with DMSO, in a volumetric flask.

6. Quality Control Standard:

6.1 2µg/mL NDMA QC Solution:

Dispense 10µL of **Quality Control Standard, 200µg/mL (4.4)** into a headspace vial, add 1.0 mL of DMSO, cap and crimp.

7. Methods of Analyses:

7.1 Procedure A - Preliminary scan*

7.1.1 2µg/mL NDMA Check Standard:

Dispense 10µL of stock standard (200µg/mL) **(5.1)** into a headspace vial, add 1.0 mL DMSO, cap and crimp.

(A higher concentration standard may be used to achieve a greater response).

7.1.2 Sample Preparation

Place 0.20-0.25g of powdered tablet or API in a headspace vial, add 1.0 mL of DMSO, cap and crimp.

7.1.3 Analyses

Perform a full mass scan on the standard **(7.1.1)** and sample **(7.1.2)** using the GC and Headspace conditions specified in **Procedure A** (Annex I).

Establish the retention time, and confirm that the mass spectrum obtained from the eluted peak in the standard solution is that of NDMA.

Continue with Quantification of NDMA, **Procedure B (7.2)**, if NDMA is detected in the sample, or with **Procedure C (LOD/LOQ determination, 7.3)** if NDMA is not detected.

*This preliminary scan method can detect NDMA at a level of 10 µg/g. As LOD/LOQ determination (Procedure C) has a lower detection limit of 0.04 µg/g, if an appreciable amount of NDMA is observed when carrying out LOD/LOQ determination (Procedure C) (7.3) the sample in question will be reanalysed using Quantification of NDMA (Procedure B).

7.2 Procedure B - Quantification of NDMA

7.2.1 Sample Preparation (Tablets)

Accurately weigh the same amount of sample (0.20-0.25g) into each of 4 Headspace vials.

7.2.1.1 Add 1.0mL of DMSO to Vial 1, cap and crimp.

7.2.1.2 Add 2 μ g (10 μ l of Stock standard solution (**5.1**)) of NDMA to Vial 2 and make up to 1.0mL with DMSO, cap and crimp.

7.2.1.3 Add 4 μ g (20 μ l of Stock standard solution (**5.1**)) of NDMA to Vial 3 and make up to 1.0mL with DMSO, cap and crimp.

7.2.1.4 Add 6 μ g (30 μ l of Stock standard solution (**5.1**)) of NDMA to Vial 4 and make up to 1.0mL with DMSO, cap and crimp.

7.2.1.5 Run all solutions using the GC and Headspace conditions described in **Procedure B** (Annex 1). Plot the standard addition curve and obtain the sample NDMA concentration (μ g/mL) from the intercept. See calculated example in Annex 2.

7.2.2 Sample Preparation (API)

7.2.2.1 Accurately weigh the same amount of sample (0.05-0.2g) into each of 4 vials.

7.2.2.2 Add 1.0mL of DMSO to Vial 1, cap and crimp.

7.2.2.3 Add 2 μ g (10 μ l of Stock standard solution (**5.1**)) of NDMA to Vial 2 and make up to 1.0mL with DMSO, cap and crimp.

7.2.2.4 Add 4 μ g (20 μ l of Stock standard solution (**5.1**)) of NDMA to Vial 3 and make up to 1.0mL with DMSO, cap and crimp.

7.2.2.5 Add 6 μ g (30 μ l of Stock standard solution (**5.1**)) of NDMA to Vial 4 and make up to 1.0mL with DMSO, cap and crimp.

7.2.2.6 Run all solutions using the GC and Headspace conditions described in **Procedure B** (Annex 1). Plot the standard addition curve and obtain the sample NDMA concentration (μ g/mL) from the intercept. See calculated example in Annex 2.

7.2.3 Include a 2.0 μ g/mL NDMA Check Standard (**7.1.1**), and a 2.0 μ g/mL NDMA QC Solution (**6.1**) with each analytical run.

The peak areas of the 2.0 μ g/mL standard and QC solution must correspond within $\pm 10\%$.

7.2.4 The correlation coefficient (r) of the calibration curve must be ≥ 0.990 .

7.3 Procedure C - LOD/LOQ determination

7.3.1 Sample Preparation (API/ Finished products)

Accurately weigh in duplicate 0.5 g of each sample (powdered) into a headspace vial.

7.3.1.1 Add 1.0mL of DMSO to Vial 1, cap and crimp.

7.3.1.2 Add 20µl (0.02µg NDMA) of 1µg/mL NDMA Standard (**5.2**), and 1.0mL of DMSO to Vial 2, cap and crimp.

7.3.1.3 Run all solutions using the conditions described in **Procedure C** (Annex 1). Ensure that a peak with a S/N of at least 3 is obtained for the 0.02µg/mL NDMA standard. Examine all chromatograms and if no NDMA peak is observed, or a peak area less than that of the standard (0.02µg/mL) is observed in the samples, results are reported as <0.04µg/g. If the sample peak area is significantly greater than that of the standard, proceed with **Procedure B (7.2)**.

Annex I – GC-MS conditions – Procedures A, B and C

Annex 2 – Standard Addition Plot



Feidhmeannacht na Seirbhíse Sláinte
Health Service Executive

Determination of NDMA (HS-GC-MS)
Method 3/30-Annex 1 GC-MS Conditions
Issue No. 1

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Procedure A

[Comment]

==== Analytical Line 1 =====

[HS-20]

Oven Temp. :120.0 °C
Sample Line Temp. :150.0 °C
Transfer Line Temp. :150.0 °C
Shaking Level :2
Multi Injection Count :1
Pressurizing Gas Pressure :80.0 kPa
Equilibrating Time. :10.00 min
Pressurizing Time. :3.00 min
Pressure Equilib. Time :0.10 min
Load Time :0.50 min
Load Equilib. Time :0.10 min
Injection Time :0.08 min
Needle Flush Time :5.00 min
GC Cycle Time :35.00 min
Check System Ready :ON
Extended System Ready Check Limit :45 min
Check GC Ready :ON
Extended GC Ready Check Limit :10 min
Analysis Mode :Constant
Needle Check :Yes
Action on Leak Check Error :Stop
Action with No Vial on Tray :Stop

[GC-2010]

Column Oven Temp. :70.0 °C
Injection Mode :Split
Flow Control Mode :Linear Velocity
Pressure :61.8 kPa
Total Flow :23.4 mL/min
Column Flow :0.97 mL/min
Linear Velocity :36.1 cm/sec
Purge Flow :3.0 mL/min
Split Ratio :20.0
High Pressure Injection :OFF
Carrier Gas Saver :OFF
Splitter Hold :OFF
Oven Temp. Program
Rate Temperature(°C) Hold Time(min)
- 70.0 12.00
3.00 100.0 2.00
60.00 240.0 2.00

< Ready Check Heat Unit >

Column Oven : Yes
HS Flow : Yes
MS : Yes
< Ready Check Detector(FTD/BID) >
< Ready Check Baseline Drift >
< Ready Check Injection Flow >
HS Flow Carrier : Yes
HS Flow Purge : Yes
< Ready Check APC Flow >
APC1 : Yes
< Ready Check Detector APC Flow >
External Wait :No
Equilibrium Time :3.0 min
< Additional Flow >
APC1 Pressure :80.0 kPa

[GC Program]

[GCMS-QP2020]

IonSourceTemp :250.00 °C
Interface Temp. :250.00 °C
Solvent Cut Time :4.00 min
Detector Gain Mode :Absolute
Detector Gain :0.93 kV
Threshold :0

[MS Table]

--Group 1 - Event 1--

Start Time :6.00min
End Time :22.00min
ACQ Mode :Scan
Event Time :0.30sec
Scan Speed :3333
Start m/z :30.00
End m/z :800.00



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Procedure B

[Comment]

==== Analytical Line 1 =====

[HS-20]
Oven Temp. :120.0 °C
Sample Line Temp. :150.0 °C
Transfer Line Temp. :150.0 °C
Shaking Level :2
Multi Injection Count :1
Pressurizing Gas Pressure :80.0 kPa
Equilibrating Time. :10.00 min
Pressurizing Time. :3.00 min
Pressure Equilib. Time :0.10 min
Load Time :0.50 min
Load Equilib. Time :0.10 min
Injection Time :0.08 min
Needle Flush Time :5.00 min
GC Cycle Time :25.00 min
Check System Ready :ON
Extended System Ready Check Limit :45 min
Check GC Ready :ON
Extended GC Ready Check Limit :10 min
Analysis Mode :Constant
Needle Check :Yes
Action on Leak Check Error :Stop
Action with No Vial on Tray :Stop

[GC-2010]
Column Oven Temp. :70.0 °C
Injection Mode :Split
Flow Control Mode :Linear Velocity
Pressure :61.8 kPa
Total Flow :23.4 mL/min
Column Flow :0.97 mL/min
Linear Velocity :36.1 cm/sec
Purge Flow :3.0 mL/min
Split Ratio :20.0
High Pressure Injection :OFF
Carrier Gas Saver :OFF
Splitter Hold :OFF
Oven Temp. Program

Rate	Temperature(°C)	Hold Time(min)
-	70.0	12.00
40.00	240.0	2.00

< Ready Check Heat Unit >
 Column Oven : Yes
 HS Flow : Yes
 MS : Yes
 < Ready Check Detector(FTD/BID) >
 < Ready Check Baseline Drift >
 < Ready Check Injection Flow >
 HS Flow Carrier : Yes
 HS Flow Purge : Yes
 < Ready Check APC Flow >
 APC1 : Yes
 < Ready Check Detector APC Flow >
 External Wait :No
 Equilibrium Time :3.0 min
 < Additional Flow >
 APC1 Pressure :80.0 kPa

[GC Program]

[GCMS-QP2020]
Micro Scan Width :0.00 amu
IonSourceTemp :250.00 °C
Interface Temp. :250.00 °C
Solvent Cut Time :4.00 min
Detector Gain Mode :Absolute
Detector Gain :0.93 kV

[MS Table]

--Group 1 - Event 1--
Start Time :8.00min
End Time :12.00min
ACQ Mode :SIM
Event Time :0.30sec
Ch1-m/z :74.00
Ch2-m/z :42.00
Ch3-m/z :43.00

Sample Inlet Unit :GC



Procedure C

==== Analytical Line 1 =====

[HS-20]
Oven Temp. :145.0 °C
Sample Line Temp. :150.0 °C
Transfer Line Temp. :150.0 °C
Shaking Level :2
Multi Injection Count :1
Pressurizing Gas Pressure :80.0 kPa
Equilibrating Time. :15.00 min
Pressurizing Time. :3.00 min
Pressure Equilib. Time :0.10 min
Load Time :0.50 min
Load Equilib. Time :0.10 min
Injection Time :0.08 min
Needle Flush Time :5.00 min
GC Cycle Time :15.00 min
Check System Ready :ON
Extended System Ready Check Limit :45 min
Check GC Ready :ON
Extended GC Ready Check Limit :10 min
Analysis Mode :Constant
Needle Check :Yes
Action on Leak Check Error :Stop
Action with No Vial on Tray :Stop

[GC-2010]
Column Oven Temp. :80.0 °C
Injection Mode :Split
Flow Control Mode :Linear Velocity
Pressure :64.9 kPa
Total Flow :13.6 mL/min
Column Flow :0.96 mL/min
Linear Velocity :36.1 cm/sec
Purge Flow :3.0 mL/min
Split Ratio :10.0
High Pressure Injection :OFF
Carrier Gas Saver :OFF
Splitter Hold :OFF
Oven Temp. Program
Rate Temperature(°C) Hold Time(min)
- 80.0 8.50
50.00 240.0 1.00

< Ready Check Heat Unit >
Column Oven : Yes
HS Flow : Yes
MS : Yes
< Ready Check Detector(FTD/BID) >
< Ready Check Baseline Drift >
< Ready Check Injection Flow >
HS Flow Carrier : Yes
HS Flow Purge : Yes
< Ready Check APC Flow >
APC1 : Yes
< Ready Check Detector APC Flow >
External Wait :No
Equilibrium Time :3.0 min
< Additional Flow >
APC1 Pressure :80.0 kPa

[GC Program]

[GCMS-QP2020]
Micro Scan Width :0.00 amu
IonSourceTemp :250.00 °C
Interface Temp. :250.00 °C
Solvent Cut Time :4.00 min
Detector Gain Mode :Absolute
Detector Gain :0.93 kV

[MS Table]

--Group 1 - Event 1--
Start Time :6.00min
End Time :9.00min
ACQ Mode :SIM
Event Time :0.30sec
Ch1-m/z :74.00
Ch2-m/z :42.00
Ch3-m/z :43.00

Sample Inlet Unit :GC

Standard Addition Plot of Peak Area Vs $\mu\text{g/mL}$ of added NDMA

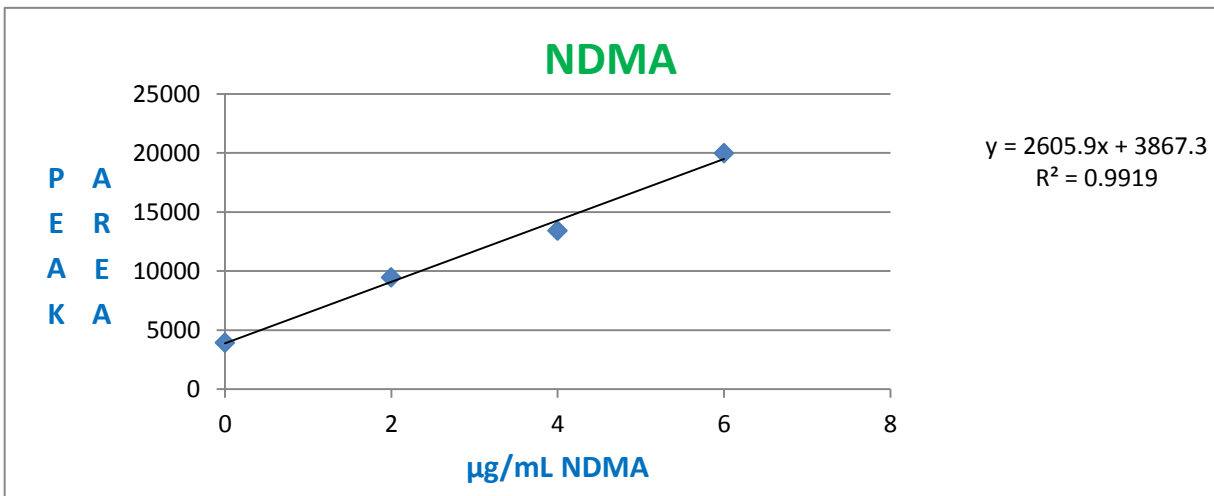
Date:

Sample

Batch:

Data Folder: GCMSsolutions\NDMA

Added NDMA ($\mu\text{g/L}$)	Area	sample (g)
0	3918	0.19908
2.0	9441	0.20165
4.0	13415	0.20004
6.0	19966	0.19946
	mean =	0.20006



Average tablets mass (g) = 0.083

API content of tablet (g) = 0.040

X Intercept = -1.4475 $\mu\text{g/mL}$

Actual value of the intercept.

➔ **7.24**

$\mu\text{g NDMA/g}$

Absolute value of the intercept divided by average mass (g) of sample used per vial.

➔ **0.60**

$\mu\text{g NDMA/tablet}$

$\mu\text{g NDMA/g}$ multiplied by average tablet mass.

➔ **15.0**

$\mu\text{g NDMA/g Valsartan}$

$\mu\text{g NDMA/tablet}$ divided by API content (g) of tablet.