Quality requirements for nanomedicines: which role for the European Pharmacopoeia?

7-8 June 2022

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mRNA as a drug: Modulating mRNA immunogenicity 300 250 200 TNF-a (pg/ml) 150 nity, Vol. 23, 165-175, August, 2005, Copyright @2005 by Elsevier Inc. DOI 10.1016/j.i muni 2005.06.008 100 Suppression of RNA Recognition by Toll-like 50 **Receptors: The Impact of Nucleoside Modification** and the Evolutionary Origin of RNA n vitro transcribed RNA-1866 R-848 tRNA total Katalin Karikó,^{1,*} Michael Buckstein,² Houping Ni,² and Drew Weissman² ¹Department of Neurosurgery poly(I):(C) total polyA+ mRNA thetic antiviral compound R-848 (Jurk et al., 24 a natural ligand has not been identified. It has been known for decades that select ipofectin nuclear cytoplasmic mitochondrial tRNA and RNA molecules have the unique property vate the immune system. It was discovered cently that secretion of interferon in response is mediated by unmethvlated CpG motifs acti ²Department of Medicine University of Pennsylvania School of Medicine Philadelphia, Pennsylvania 19104 *E. coli* RNA mammalian RNA





How to get mRNA into cells?

Proc. Natl. Acad. Sci. USA Vol. 86, pp. 6077–6081, August 1989 Biochemistry

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Fig. 4. RNA transfection in a variety of cell types. Fifty micrograms of lipofectin liposomes was used to transfect various cell lines either with (+) or without (-) 20 μg of mRNA (Cap-βg Luc βg An). Lysates were prepared 8 hr after addition of lipofectin and were analyzed for luciferase specific activity as before.



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able1: Quality Attributes for mRNA Drug Substance					
Quality	Attribute	Method			
Identity	Sequence confirmation	Next generation sequencing (NGS)			
		Sanger sequencing			
		Reverse Transcriptase – PCR			
Content	RNA content	RT-qPCR and RT-dPCR, Ultraviolet Spectroscopy			
Integrity	Percentage of Intact mRNA and fragment mRNA	Capillary gel electrophoresis			
	5" cap	IP-RP-HPLC			
	3' poly(A)	RP-HPLC			
	mRNA Integrity	Gel electrophoresis			
5	Product related impurities - dsRNA	Immunoblot			
Purity	Residual DNA template	qPCR			
	Endotoxin	USP <85>			
Safety	Bioburden	USP <61>, <62>, <1115>			
	Sterility	USP <71>			
Other	Appearance	USP <1>, <790>			
Other	рН	USP <791>			











- Industry gold standard techniques (CE and SEC) could not detect this impurity
- Both NGS and oligonucleotide mapping showed an identical profile between the MP and LP, suggesting very low abundance of non-site-specific modifications.
- Nucleoside profiling revealed several abundant mass-to-charge (m/z) values that were exclusively found in the isolated LP.
- MS/MS revealed that the unique m/z are lipid-modified nucleosides (modified on the nucleobase).

1	 Reaction modeling studies was performed to identify lipid contributors to adduct formation (data not shown)- electrophilic/oxidative impurities originating from the ionizable cationic lipid were identified as reactants, resulting in adduct formation 			
		Source: A novel mechanism for the loss of mRNA activity Packer M, et al. Nat Commun. 2021.	ty in lipid nanoparticle delivery systems.	
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Genomic Medicines are the Future



































Analytical Requirements to Assess RNA-LNPs





















Analytical Methods Overview								
Test	Method	Release Testing	In Process Testing					
Appearance	Visual Inspection: USP <790> / Ph.Eur. 2.9.20	Х						
pН	Potentiometric: USP <791> / Ph.Eur 2.2.3	Х	(X)*					
Osmolality	Freezing Point Depression: USP <785> / Ph.Eur. 2.2.35	Х	(X)*					
Bacterial Endotoxins	USP<85> / Ph.Eur 2.6.14.	Х						
Sterility/BioBurden	USP<71> / Ph.Eur 2.6.1.	Х	(X)*					
Particulate Matter	USP<788> / Ph.Eur 2.9.19.	Х						
Elemental Impurities	USP<233> / Ph.Eur 2.4.20	Х						
RNA Identity/Integrity	Capillary Electrophoresis or Bioanalyzer	Х	X**					
Particle Size/PDI	Dynamic Light Scattering	Х	X**					
RNA Content/Encapsulation	Ribogreen Assay	Х	X**					
Lipid Content	UPLC-CAD	Х	X**					
Lipid:RNA Ratio	Calculation	Х						
Potency Bioassay	In Vitro Assay	Х						
*applied for all aqueous buffer systems **applied for all LNP processing steps								
	Methods Over Test Appearance pH Osmolality Bacterial Endotoxins Sterility/BioBurden Particulate Matter Elemental Impurities RNA Identity/Integrity Particle Size/PDI RNA Content/Encapsulation Lipid Content Lipid:RNA Ratio Potency Bioassay s buffer systems **applied for	MethodTestMethodAppearanceVisual Inspection: USP <790> / Ph.Eur. 2.9.20pHPotentiometric: USP <791> / Ph.Eur 2.2.3OsmolalityFreezing Point Depression: USP <785> / Ph.Eur. 2.2.35Bacterial EndotoxinsUSP <85> / Ph.Eur 2.6.14.Sterility/BioBurdenUSP <71> / Ph.Eur 2.6.1.Particulate MatterUSP <788> / Ph.Eur 2.9.19.Elemental ImpuritiesUSP <233> / Ph.Eur 2.4.20RNA Identity/IntegrityCapillary Electrophoresis or BioanalyzerParticle Size/PDIDynamic Light ScatteringRNA Content/EncapsulationUPLC-CADLipid ContentUPLC-CADLipid ContentIn Vitro Assaysbuffer systems **applied turb rocessing steps	Methods Overview Method Release Testing Appearance Visual Inspection: USP <790> / Ph.Eur. 2.9.20 X PH Potentiometric: USP <791> / Ph.Eur 2.9.20 X Osmolality Freezing Point Depression: USP <785> / Ph.Eur 2.2.35 X Bacterial Endotoxins USP <85> / Ph.Eur 2.6.14. X Sterility/BioBurden USP <785> / Ph.Eur 2.6.1. X Particulate Matter USP <788> / Ph.Eur 2.9.19. X Elemental Impurities USP <233> / Ph.Eur 2.4.20 X RNA Identity/Integrity Capillary Electrophoresis or Bioanalyzer X RNA Ribogreen Assay X Lipid Content UPLC-CAD X Lipid Content UPLC-CAD X Potency Bioassay In Vitro Assay X Suffer systems **applied tor all LNP processing steps X X					

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